

## Thyroid Hormone Regulation of Na,K-ATPase Subunit-mRNA Expression in Neonatal Rat Myocardium

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**Summary.** Regulation of Na,K-ATPase mRNA $_{\alpha}$  isoform and mRNA $_{\beta}$  expression by thyroid hormone (T<sub>3</sub>) in neonatal rat myocardium was examined. In euthyroid neonates between ages of 2 and 5 days, mRNA $_{\alpha 1}$ , mRNA $_{\alpha 3}$ , and mRNA $_{\beta 1}$  abundances were nearly constant while mRNA $_{\alpha 2}$  was undetectable. During the interval between postnatal days 5 and 15, mRNA $_{\alpha 3}$  decreased to negligible levels and mRNA $_{\alpha 2}$  became expressed and increased in abundance to account for ~20% of the mRNA $_{\alpha}$  pool by the 15<sup>th</sup> postnatal day. To examine the effect of T<sub>3</sub> on this developmental program, neonates were injected with 75  $\mu$ g T<sub>3</sub>/100 g body weight or diluent alone on the second and third postnatal days and myocardial Na,K-ATPase subunit-mRNA abundances were determined on the third and fourth postnatal days. Because T<sub>3</sub> treatment increased the RNA/DNA ratios of myocardial tissue, the subunit-mRNA abundances were normalized per unit DNA. Following 24 and 48 hr of T<sub>3</sub> treatment, the abundances of mRNA $_{\alpha 1}$ , mRNA $_{\alpha 3}$ , and mRNA $_{\beta 1}$  increased, while mRNA $_{\alpha 2}$  continued to remain undetectable during the 2-day interval between the second to fourth postnatal days. It is concluded that T<sub>3</sub> augments the abundance of Na,K-ATPase subunit mRNAs that are already being expressed in the neonatal rat myocardium. The results further suggest that T<sub>3</sub> does not act as a "molecular switch" in the developmental expression of the mRNA $_{\alpha}$  isoforms in rat myocardium during the first four postnatal days.

**Key Words** T<sub>3</sub> · Na,K-ATPase mRNA isoforms · developmental regulation of Na,K-ATPase

### Introduction

Na<sup>+</sup>,K<sup>+</sup>-activated adenosine triphosphatase (Na,K-ATPase; Na,K-pump) is an integral plasma membrane enzyme through whose function intracellular Na<sup>+</sup> and K<sup>+</sup> concentrations are maintained [21]. The enzyme consists of two dissimilar subunits,  $\alpha$  and  $\beta$ , that are present in equimolar ratio in purified preparations isolated from a variety of tissues [21]. The  $\alpha$ , or "catalytic," subunit of Na,K-ATPase has an  $M_r$  of ~110,000 and is present as three isoforms designated  $\alpha_1$ ,  $\alpha_2$ , and  $\alpha_3$  that are expressed in a tissue-specific manner [8, 29, 34, 35, 43, 46]. The  $\beta$

subunit of Na,K-ATPase has a  $M_r$  of ~55,000 and is heavily glycosylated [21, 36]. Recently a putative isoform of the  $\beta$  subunit mRNA (designated  $\beta_2$ ) has been described, and the corresponding polypeptide is expressed in rat brain but not in rat myocardium [26, 38]. Isoforms of Na,K-ATPase exhibit different sensitivities towards cardiac glycosides such as ouabain; the  $\alpha_1$  isoform of the rat is least sensitive with a  $K_I$  of  $\sim 10^{-4}$  M for ouabain, while both the rat  $\alpha_2$  and  $\alpha_3$  isoforms exhibit  $K_I$ 's of  $\sim 10^{-7}$  M for ouabain [1, 11, 14–16, 25, 34, 40, 41, 44]. Rat kidney and liver express the  $\alpha_1$  isoform almost exclusively, while both  $\alpha_1$  and  $\alpha_2$  isoforms are expressed in cardiac and skeletal muscle [8, 11, 16, 18, 29, 41, 46]. All three  $\alpha$ -isoforms are expressed in rat cerebral cortex [8, 29, 34, 35, 43, 46]. In a recent study on the developmental expression of Na,K-ATPase subunit mRNAs in several rat tissues, Orłowski and Lingrel [29] reported that mRNA $_{\alpha 2}$  is not expressed in the myocardium at birth, while mRNA $_{\alpha 1}$  and lesser quantities of mRNA $_{\alpha 3}$  are present. By the 10<sup>th</sup> to 14<sup>th</sup> day of postnatal life, however, mRNA $_{\alpha 3}$  virtually disappears, while mRNA $_{\alpha 2}$  becomes detectable between the fifth to seventh postnatal days and reaches near-adult levels by day 28 at which time it remains a minor component of myocardial mRNA $_{\alpha}$  pool.

Triiodothyronine (T<sub>3</sub>) stimulates Na,K-ATPase activity in a variety of mammalian target tissues including the myocardium [2, 4, 10, 11, 14, 17, 18, 22, 23, 27, 32]. The enhancement of enzyme activity is attributable to an increase in the number of enzyme units through increased biosynthesis of the enzyme [2, 22–24]. T<sub>3</sub> augments the abundance of the mRNAs encoding the subunits of the enzyme in several target tissues including myocardium, an effect that precedes the stimulation of Na,K-ATPase activity and in part results from enhanced transcription of the corresponding genes [2, 9–11, 18, 27]. We have recently reported that while the  $\alpha_2$ -isoform and

mRNA<sub>α2</sub> are minor components of adult rat myocardial Na,K-ATPase and mRNA<sub>α</sub> pools, respectively, the abundance of mRNA<sub>α2</sub> and high-ouabain-sensitivity enzyme activity are preferentially augmented by T<sub>3</sub> [11]. A near-selective T<sub>3</sub>-induced increase in the high-ouabain-sensitivity form of the enzyme has additionally been reported in adult rat diaphragm [14].

In view of the transcriptional regulation of Na,K-ATPase by T<sub>3</sub>, the preferential enhancement of mRNA<sub>α2</sub> and α<sub>2</sub>-isoform by T<sub>3</sub> in rat myocardium and skeletal muscle [10, 11, 14, 18], and the developmental program of Na,K-ATPase mRNA<sub>α</sub>-isoform expression in the myocardium during the early postnatal period [29], we examined the possibility that the "molecular switching" between mRNA<sub>α2</sub> and mRNA<sub>α3</sub> is mediated by thyroid hormone. This possibility was further suggested by the observations that thyroid hormone concentrations in rat plasma at birth are negligible, and that T<sub>4</sub> and T<sub>3</sub> concentrations increase during the first few days postnatally to reach  $\frac{1}{3}$  to  $\frac{1}{2}$  adult concentrations by the seventh to tenth day [6, 7, 45], an interval during which the aforementioned changes in Na,K-ATPase mRNA<sub>α</sub>-isoform expression are manifested. The hypothesis was also consistent with the finding that T<sub>3</sub> can act as a "molecular switch" in the expression of the myosin heavy chain isoforms [12, 13, 19, 33]. In the present study we document the effects of T<sub>3</sub> on Na,K-ATPase mRNA<sub>α</sub>-isoform and mRNA<sub>β1</sub> expression in euthyroid neonatal rat myocardium during the first four postnatal days. We find that T<sub>3</sub> augments the abundance of the subunit mRNAs that are already being expressed in this tissue, and that the hormone does not appear to act as a "molecular switch" in myocardial Na,K-ATPase mRNA<sub>α</sub>-isoform expression.

## Materials and Methods

### MATERIALS

Standard compounds including 3,3',5-triiodothyronine (T<sub>3</sub>) were obtained from Sigma Chemical. <sup>32</sup>P-α-TTP (3000 Ci/mmol) was purchased from Amersham. Nitrocellulose paper (BA-85) and "nick-translation" kits were obtained from Schleicher and Schuell and Bethesda Research Laboratories, respectively.

### METHODS

#### *Animals and Preparation of Tissues*

Litters were obtained on the first postnatal day and the mothers maintained on regular rat chow. In experiments designed to study the expression of Na,K-ATPase mRNA<sub>α</sub> isoforms during devel-

opment, noninjected neonates were studied up to 15 days of age. In experiments examining the effects of thyroid hormone, each litter was randomly assigned to diluent or T<sub>3</sub> groups and kept in separate cages. Beginning on the second postnatal day, neonates were injected subcutaneously with 75 μg T<sub>3</sub>/100 g body weight [28] dissolved in 200 μl of 1 mM NaOH solution or diluent alone once daily and were sacrificed on postnatal days 3 and 4. Hearts were rapidly removed and frozen in liquid nitrogen and stored for less than two weeks prior to isolation of RNA.

#### *Quantitation of Tissue Protein, RNA, and DNA*

Three to five hearts were pooled at each time point and for each experimental condition. Methods previously described were employed without modification [2, 10].

#### *Isolation of RNA and Analysis of Na,K-ATPase Subunit-mRNA Abundances*

In each experiment 4–6 hearts were pooled for the isolation of RNA at each time point from the T<sub>3</sub>- and diluent-treated neonates. Total RNA was isolated by ultracentrifugation as described by Chirgwin et al. [3]. Quantitation of RNA, electrophoresis, blotting, hybridization, and washing conditions were as described previously [2, 11, 42]. Approximately 20 μg of total RNA isolated from each of the different treatment groups was loaded per lane in four replicate blots were hybridized with plasmids containing full-length cDNA<sub>α1</sub>, cDNA<sub>α2</sub>, cDNA<sub>α3</sub>, and cDNA<sub>β1</sub> "nick-translated" to near-equivalent specific activity ( $\sim 5 \times 10^8$  cpm/μg DNA;  $\sim 3 \times 10^7$  cpm used per blot) [35, 36]. Following autoradiography, the intensity of the corresponding bands were determined by densitometry [2]. Ethidium-bromide staining of the ribosomal RNA bands was monitored on the gels and on the nitrocellulose papers to ensure equal RNA loading of the gels and complete transfer of the RNA [9, 11, 47].

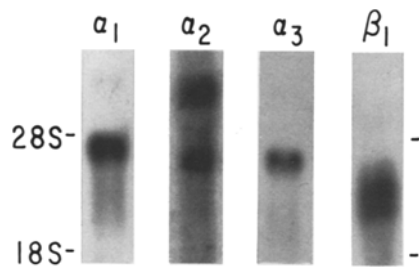
To measure the relative abundances of Na,K-ATPase subunit mRNAs, autoradiograms were prepared at various intervals to ensure that the signals were in the linear response range of the film. The resulting density readings were corrected for the elapsed time of exposure, length of the cDNAs [35, 36], and the amount of radioactivity used in each hybridization reaction.

### STATISTICAL ANALYSIS

Data from the different independent experiments were averaged and are presented as means ± SE. Analysis of variance and least-square difference between the means was used throughout to test for statistical significance [39]. *P* values less than 0.05 were taken as significant.

## Results

A Northern blot demonstrating the expression of mRNA<sub>α1</sub>, mRNA<sub>α3</sub>, and mRNA<sub>β1</sub> in 2-day-old and mRNA<sub>α2</sub> in 15-day-old euthyroid rat myocardial tis-



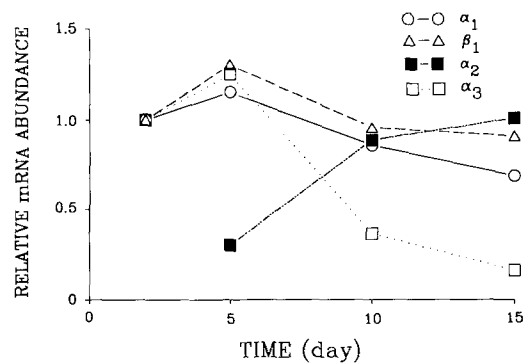
**Fig. 1.** Expression of Na,K-ATPase  $\alpha_1$ -,  $\alpha_3$ -, and  $\beta$ -mRNA in 2-day-old and of  $\alpha_2$ -mRNA in 15-day-old euthyroid rat myocardium. RNA was isolated from 4–6 pooled neonatal rat hearts on the second or fifteenth postnatal days. Twenty  $\mu\text{g}$  of total RNA was fractionated, blotted, and probed with Na,K-ATPase cDNAs. Blots probed with cDNA $_{\alpha_1}$ , cDNA $_{\alpha_3}$ , and cDNA $_{\beta_1}$  were from 2-day-old neonates. Note that the myocardial RNA probed with cDNA $_{\alpha_2}$  was obtained from 15-day-old neonates. The positions of the 28 S and 18 S ribosomal-RNA bands are shown

**Table 1.** Relative abundances of Na,K-ATPase mRNA $_{\alpha}$  isoforms and mRNA $_{\beta_1}$  in myocardium of euthyroid neonatal rats on the second postnatal day

mRNA	Relative molar abundance
mRNA $_{\alpha_1}$	1.0 $\pm$ 0.1
mRNA $_{\alpha_2}$	“undetectable”
mRNA $_{\alpha_3}$	0.4 $\pm$ 0.1
mRNA $_{\beta_1}$	1.9 $\pm$ 0.1

Three samples of RNA were isolated from pooled myocardial tissue of 2-day-old euthyroid neonates from three separate litters. Twenty  $\mu\text{g}$  of total RNA obtained from each of the pooled myocardial samples were fractionated in 1% agarose gels in four replicate gels and following blotting were probed with cDNA $_{\alpha_1}$ , cDNA $_{\alpha_2}$ , cDNA $_{\alpha_3}$ , and cDNA $_{\beta_1}$ , respectively. The intensities of the resulting bands on the autoradiograms, corrected for the specific activity of the respective probes, the length of the cDNAs, and the period of exposure, were normalized to the mean intensity obtained utilizing the cDNA $_{\alpha_1}$  probe. The SE of mRNA $_{\alpha_1}$  value reflects the variance of the abundances in the three RNA samples. mRNA $_{\alpha_2}$  abundance was below the limit of reliable detection. The results are expressed as means  $\pm$  SE ( $n = 3$ ).

sue is shown in Fig. 1. In the figure, RNA obtained from 15-day-old rats was used for mRNA $_{\alpha_2}$  analysis because, as will be noted below, this mRNA species is not detectable in 2-day-old rat myocardium. The positions of the mRNA bands are in accordance to previous reports [11, 29, 35, 36, 46]. The relative abundances of  $\alpha_1$ -,  $\alpha_2$ -,  $\alpha_3$ -, and  $\beta_1$ -mRNA in myocardium of 2-day-old euthyroid neonates were then determined. Replicate blots prepared from RNA isolated from three different litters were probed with the corresponding cDNAs and the relative density readings of autoradiograms determined (Table 1).



**Fig. 2.** Relative abundance of Na,K-ATPase subunit-mRNA isoforms in 2-day-old to 15-day-old euthyroid neonatal rat myocardium. RNA was isolated from 3–6 pooled neonatal rat hearts on the days indicated. Approximately 10  $\mu\text{g}$  of total RNA from each time point was fractionated in adjacent lanes in four replicate blots and probed with full-length cDNA $_{\alpha_1}$ , cDNA $_{\alpha_2}$ , cDNA $_{\alpha_3}$ , and cDNA $_{\beta_1}$ . The experiment was repeated four times and the results averaged. To express the relative abundances of the mRNAs per unit DNA at each of the time points, the abundances were multiplied by the RNA/DNA ratio at the corresponding time points; RNA/DNA ratios were 0.69, 0.77, 1.04, and 0.83 on days 2, 5, 10, and 15, respectively. The resulting mRNA abundances were then normalized to their corresponding day-2 values except for mRNA $_{\alpha_2}$  abundance on day 15 which was set to 1.0. mRNA $_{\alpha_2}$  abundance on day 2 was low and could not be reliably determined. None of the changes in mRNA $_{\alpha_1}$  and mRNA $_{\beta_1}$  abundances were significant. mRNA $_{\alpha_2}$  abundance on day 5 was significantly lower than its value on days 10 and 15, while mRNA $_{\alpha_3}$  abundances on days 10 and 15 were significantly lower than the values on days 2 and 5

On the second postnatal day, mRNA $_{\alpha_1}$  abundance exceeded mRNA $_{\alpha_3}$  abundance by 2.5-fold and mRNA $_{\alpha_2}$  was undetectable; mRNA $_{\beta_1}$  abundance exceeded the total mRNA $_{\alpha}$  pool by  $\sim$ 1.4-fold. On prolonged exposure (5–7 days) bands corresponding to mRNA $_{\alpha_2}$  became faintly visible but could not be quantified because of the increase in background exposure.

We next examined the expression of Na,K-ATPase subunit mRNAs during neonatal development of euthyroid rats between the second to fifteenth postnatal days (Fig. 2). In the figure, subunit-mRNA abundances at each time point were standardized per unit DNA and, for purposes of comparison, the abundance of each of  $\alpha_1$ -,  $\alpha_3$ -, and  $\beta_1$ -mRNA on the second postnatal day and that of mRNA $_{\alpha_2}$  on the 15<sup>th</sup> postnatal day were set to 1.0. The abundance of mRNA $_{\beta_1}$  did not change significantly during the 13-day interval. mRNA $_{\alpha_1}$  abundance decreased to a value of 0.65 by day 15, but the change was not significant ( $P = 0.10$ ). mRNA $_{\alpha_2}$  became measurable on day 5 and increased in abundance such that by day 15 mRNA $_{\alpha_2}$ /mRNA $_{\alpha_1}$  ratio was 0.25. In contrast to mRNA $_{\alpha_2}$ , mRNA $_{\alpha_3}$  abundance was significantly

**Table 2.** Effect of T<sub>3</sub> on the RNA, DNA, and protein content of neonatal rat myocardium between the second and fourth postnatal days

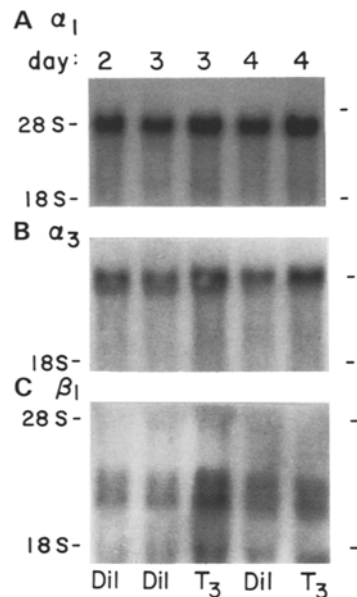
Age (day)	T <sub>3</sub>	RNA (mg/g)	DNA (mg/g)	Protein (mg/g)	RNA/DNA (mg/mg)
2	-	4.5 ± 0.5	6.7 ± 0.7	125 ± 9	0.68 ± .03
3	-	3.8 ± 0.1	5.2 ± 0.3	107 ± 3	0.73 ± .03
3	+	4.6 ± 0.4	5.6 ± 0.4	118 ± 4	0.81 ± .06
4	-	4.2 ± 0.2	6.9 ± 0.4	134 ± 6	0.61 ± .02
4	+	5.6 ± 0.6	6.8 ± 0.8	142 ± 10	0.83 ± .06 <sup>a</sup>

Beginning on the second postnatal day, rats were treated daily with T<sub>3</sub> (75 µg/100 g body weight) or diluent alone and sacrificed on the postnatal days indicated. Hearts from 3–5 rats were pooled to obtain a single sample for analysis from each of the treatment groups shown. Results of tissue contents are expressed as mg/g wet weight (mean ± SE; n = 4).

<sup>a</sup> Denotes P < 0.05.

decreased on day 10 and reached ~15% of its day-2 value by the 15<sup>th</sup> postnatal day. These data are consistent with the results reported previously by Orłowski and Lingrel [29], and demonstrate that α<sub>1</sub>-, α<sub>3</sub>-, and β<sub>1</sub>-mRNA abundances are virtually constant during the period between the second to the fifth postnatal days, and that mRNA<sub>α2</sub> becomes detectable by the fifth postnatal day.

To examine whether T<sub>3</sub> increases the abundance of Na,K-ATPase subunit mRNAs and elicits a premature “switching” of the Na,K-ATPase mRNA<sub>α</sub> expression in the myocardium, the effect of T<sub>3</sub> on the abundance of subunit mRNAs was determined between the second and fourth postnatal days. Because of potential changes in tissue RNA content resulting from T<sub>3</sub> treatment [2, 10, 11], we first measured myocardial protein, RNA, and DNA content in diluent- and T<sub>3</sub>-treated neonates (Table 2). T<sub>3</sub> increased the RNA content and the RNA/DNA ratio of the myocardium on the third and fourth postnatal days, but only the change in RNA/DNA ratio on day 4 was statistically significant. The effect of T<sub>3</sub> on Na,K-ATPase subunit-mRNA abundances is shown in Fig. 3. The abundances of mRNA<sub>α1</sub>, mRNA<sub>α3</sub>, and mRNA<sub>β1</sub> appeared to be increased by T<sub>3</sub>. This experiment was repeated four times and the averaged results are shown in Fig. 4. In the figure, all values have been standardized per unit DNA based upon the values listed in Table 2. The abundance of the three mRNAs remained unchanged in the diluent-treated neonates, in accordance to the results of Fig. 2. In contrast, T<sub>3</sub> increased the abundance of the three subunit mRNAs significantly both at 24 and 48 hr. The abundance of mRNA<sub>β1</sub> showed a larger increment than that of the α-subunit mRNAs at 24 hr, and its abundance then declined to reach a

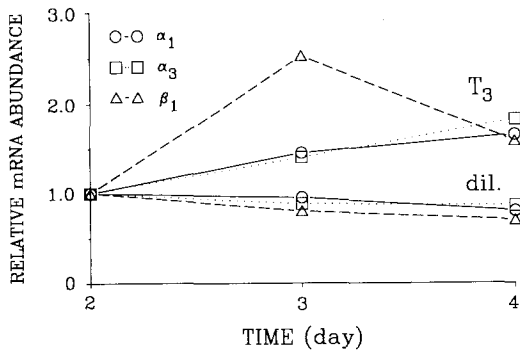


**Fig. 3.** Effect of T<sub>3</sub> on the expression of myocardial Na,K-ATPase mRNA subunit mRNAs between the second to fourth postnatal days. Litters were randomly assigned to two treatment groups and the neonates received either T<sub>3</sub> (75 µg/100 g body weight) or diluent alone on postnatal days 2 and 3 and were studied on postnatal days 3 and 4. RNA was isolated from 4–6 pooled hearts, fractionated, and probed with the Na, K-ATPase subunit cDNAs as described in the legend to Fig. 1. Panels A, B, and C show the results of the corresponding blots probed with cDNA<sub>α1</sub>, cDNA<sub>α3</sub>, and cDNA<sub>β1</sub>, respectively

net increment comparable to the increments in α<sub>1</sub>- and α<sub>3</sub>-mRNAs by 48 hr. T<sub>3</sub> did not result in a premature disappearance mRNA<sub>α3</sub>, and indeed mRNA<sub>α3</sub> abundance increased following treatment with the hormone. mRNA<sub>α2</sub> abundance was too low to be accurately measured during the 48-hr interval and treatment with T<sub>3</sub> did not appear to induce a premature expression of the mRNA<sub>α2</sub> isoform.

## Discussion

The developmental changes in rat myocardial Na,K-ATPase mRNA abundances shown in Figs. 1 and 2 are consistent with the results reported by Orłowski and Lingrel [29] and demonstrate that, during the short interval between the second to the fifth postnatal days, the abundances of mRNA<sub>α1</sub>, mRNA<sub>α3</sub>, and mRNA<sub>β</sub> remain relatively constant. The results furthermore show that the abundance of mRNA<sub>α3</sub>



**Fig. 4.** Effect on T<sub>3</sub> on the relative abundance of Na,K-ATPase subunit mRNAs between the second to fourth postnatal days. Conditions were as described in the legend to Fig. 3. The experiment was repeated four times and the results averaged. To express the relative abundances of the mRNAs per unit DNA at each of the time points and for each treatment condition, the respective abundances were multiplied by the RNA/DNA ratios listed in Table 2. The resulting mRNA abundances were then normalized to their corresponding day-2 values. None of the changes in the mRNA abundances in the diluent-treated group were significant. In contrast, all the increments in the T<sub>3</sub>-treated group on days 3 and 4 were significant. The abundance of mRNA<sub>β1</sub> in T<sub>3</sub>-treated neonates on day 3 was significantly larger than its value on day 4

markedly decreases and that of mRNA<sub>α2</sub> increases between the 5<sup>th</sup> and 15<sup>th</sup> postnatal days, as reported previously [29].

Previous studies have demonstrated that thyroid hormone concentrations in the serum of neonatal rats are negligible at birth and increase dramatically during the first two weeks postnatally [6, 7, 45]. Furthermore, thyroid hormones are known to augment the activity of Na,K-ATPase in target tissues including myocardium, an enhancement that is preceded by increases in the abundances of the subunit mRNAs that in turn are mediated in part at the transcriptional level [2, 10, 11, 27]. We thus hypothesized that T<sub>3</sub> may act as a “molecular switch” during the early postnatal development of myocardial Na,K-ATPase expression. We elected to examine the effect of T<sub>3</sub> on the expression of Na,K-ATPase subunit mRNAs between days 2 and 4 postnatally, with the prediction that if T<sub>3</sub> action is instrumental in the mRNA<sub>α</sub>-isoform switching, then a premature expression of mRNA<sub>α2</sub> and disappearance of mRNA<sub>α3</sub> should be observed following T<sub>3</sub> treatment. In experiments designed to test this hypothesis we found that T<sub>3</sub> treatment increased the abundances of α<sub>1</sub>-, α<sub>3</sub>-, and β<sub>1</sub>-mRNAs in the myocardium, demonstrating that Na,K-ATPase subunit-mRNA expression in the neonatal rat heart is regulated by T<sub>3</sub> (Figs. 3 and 4). Treatment with T<sub>3</sub> did not result in a premature induction of the mRNA<sub>α2</sub> isoform. The abun-

dance of mRNA<sub>α3</sub>, however, increased significantly with T<sub>3</sub>, a finding that has not been described previously. These results suggest that the hormone does not act as a “molecular switch” in the respective “up”- and “down”-regulation of myocardial Na, K-ATPase mRNA<sub>α2</sub> and mRNA<sub>α3</sub> expression. Instead the findings demonstrate that T<sub>3</sub> enhances the expression of the Na,K-ATPase subunit genes that are already being expressed in this tissue at the time of treatment and suggest that the switching between myocardial α<sub>2</sub>- and α<sub>3</sub>-mRNA isoforms is regulated by mechanisms other than thyroid hormone action. The above inference is consistent with the lack of mRNA<sub>α3</sub> induction in myocardium of adult hypothyroid rats treated with “receptor-saturating” doses of T<sub>3</sub> [11, 28]. It should be noted that in the present study the effect of T<sub>3</sub> was examined during the interval between the second and fourth postnatal days. Thus the possibility that the hormone may still play an important role in Na,K-ATPase mRNA<sub>α</sub> switching at later developmental stages cannot be excluded. In a recent study Orłowski and Lingrel [30] examined the effects of T<sub>3</sub> and dexamethasone on the expression of Na,K-ATPase mRNA<sub>α</sub> isoforms in rat neonatal cardiac myocytes in culture. They too found that the addition of T<sub>3</sub> in this *in-vitro* system did not elicit a “switching” between the mRNA<sub>α</sub> isoforms. However, a complex relationship between mRNA<sub>α</sub>-isoform expression following the addition of dexamethasone was noted, suggesting a potential role of this latter hormone during myocardial development [30].

The results of the present study demonstrating a lack of T<sub>3</sub> effect on “switching” of Na,K-ATPase mRNA<sub>α</sub> isoforms should be contrasted to previous results demonstrating that T<sub>3</sub> acts as a “molecular switch” in the regulation of skeletal muscle and cardiocyte myosin heavy chain expression [12, 13, 19, 20, 33]. This difference might reflect the influence of specific regulatory mechanisms operative in the control of myosin heavy chain and Na,K-ATPase gene expression. Consistent with the above interpretation is the presence of “thyroid response elements” upstream to the α- and β-myosin heavy chain genes and the lack of such an element in the 5′-upstream flanking region of the human Na,K-ATPase α<sub>2</sub> gene [12, 13, 19, 37]. Although it is not known whether “thyroid response elements” are present upstream in the other Na,K-ATPase subunit genes, and T<sub>3</sub> regulatory sites may be located at sites other than 5′-upstream to the α<sub>2</sub> gene, the above findings raise the possibility of indirect regulation of Na,K-ATPase gene expression by T<sub>3</sub>. Indeed, we have noted that thyroidal enhancement of mRNA<sub>β1</sub> abundance in Clone 9 cells is abrogated by prior exposure to cycloheximide, while ongoing protein

synthesis is not required for the induction of mRNA<sub>α1</sub> by the hormone [9]. Alternatively, it should be noted that adrenergic innervation and catecholamine uptake capacity of sympathetic neurons in neonatal rat myocardium is poorly developed at birth and matures during the first several days of postnatal development [5, 31]. It is thus possible that events associated with adrenergic neuronal activity may influence the pattern of myocardial Na,K-ATPase mRNA<sub>α</sub>-isoform expression. Further studies are thus required to elucidate mechanisms underlying the molecular "switching" of myocardial Na,K-ATPase expression, and to explore the basis of responsiveness of these genes to T<sub>3</sub> at various stages of the developmental program.

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